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## Synthesis of the tetrasaccharide side chain of the major glycoprotein of the *Bacillus anthracis* exosporium

Rina Saksena, Roberto Adamo and Pavol Kováč\*

NIDDK, LMC, National Institutes of Health, Bethesda, MD 20892-0815, USA

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Abstract—An α-glycoside of the tetrasaccharide sequence β-Ant- $(1 \rightarrow 3)$ -α-L-Rhap- $(1 \rightarrow 3)$ -α-L-Rhap- $(1 \rightarrow 2)$ -α-L-Rhap whose aglycon allows conjugation to suitable carriers was synthesized. The NMR characteristics of the compound are virtually identical with those of the α-anomer of the tetrasaccharide isolated from the major glycoprotein of the *Bacillus anthracis* exosporium. Thus, the correct structure of the natural product has been proven by chemical synthesis. © 2005 Elsevier Ltd. All rights reserved.

Bacillus anthracis is the causative agent of anthrax. Because of concerns regarding the use of some of its forms as a biological weapon, there is a worldwide effort to develop a potent vaccine for the disease. Since B. anthracis is a spore-forming pathogen, a strategy toward a vaccine for anthrax could be based on targeting spores with antibodies that are specific for the tetrasaccharide  $\beta$ -Ant-(1  $\rightarrow$  3)-α-L-Rhap-(1  $\rightarrow$  3)-α-L-Rhap-(1  $\rightarrow$  2)-L-Rhap (1), which was recently discovered<sup>2</sup> as the oligosaccharide side chain of the collagen-like region of the major glycoprotein of the pathogen's exosporium. Within our work toward a conjugate vaccine for anthrax we have prepared and here report a synthesis (Scheme 1) of sequence 1 in the form of the  $\alpha$ -glycoside 20 whose aglycone can be further transformed to allow conjugation to suitable carriers.

The initial glycosyl acceptor  $4^{13}$  was synthesized by reaction of the known<sup>3</sup> thioglycoside **2** with methyl 6-hydroxyhexanoate<sup>4</sup> ( $\rightarrow$ 3) followed by deacetylation.

Extension of the oligosaccharide chain to give rhamnotrioside 9 was effected by glycosylation of 4 with the disaccharide building block 8. To prepare 8, the known<sup>5</sup> thioglycoside 6 was treated with glycosyl chloride 7, which was obtained by treatment of 5<sup>5</sup> with chlorine.<sup>6</sup>

Keywords: Anthrax; Anthrose; Exosporium glycoprotein; Thioglycoside; Oligosaccharide.

Subsequent deacetylation of 9 gave the trisaccharide glycosyl acceptor 10.

Synthesis of **20** required formation of 1,2-trans glycosidic linkage, which could be problematic with a glycosyl donor bearing a non-participating group at O-2, such as the 2-Omethyl group in anthrose. Therefore, the versatile 2-Obromoacetylated glycosyl donor 14 (Scheme 2) was prepared. In addition to being a participating group, the bromoacetyl function<sup>7,8</sup> can be selectively removed in the presence of other acyl groups. Thus, donor 14 can be used in alternative syntheses of 1 involving acylprotected intermediates. Compound 14 was obtained as shown in Scheme 2. Accordingly, methyl 4-azido-3-O-benzyl-4,6-dideoxy-α-p-glucopyranoside<sup>9</sup> (11) was bromoacetylated, 10 and the formed 12 was acetolyzed to give 13. The anomeric mixture of acetates thus obtained was treated with EtSH under Lewis acid catalysis to give 14. Condensation of the trirhamnoside glycosyl acceptor 10 with 14 gave the fully protected tetrasaccharide 15, which was sequentially de-O-bromoacetylated (-16), methylated<sup>11</sup> ( $\rightarrow$ **17**), and treated with H<sub>2</sub>S,<sup>12</sup> to selectively reduce the azido group to amino function, to afford 18. Treatment of 18 with 3-hydroxy-3-methylbutyric acid in the presence of HATU {N-[(dimethylamino)-1H-1,2,3triazolo[4,5-b]pyridin-1-ylmethylene]-N-methylmethanaminium hexafluorophosphate N-oxide} gave butanamido derivative 19, which was fully deprotected by hydrogenolytic debenzylation, to give the target tetrasaccharide 20 equipped with a spacer that makes it amenable for conjugation to proteins or other suitable carriers. NMR data observed for **20** in DMSO- $d_6$  (Tables 1 and 2) are virtually identical to those reported<sup>2</sup> for the solution of  $\alpha$ -anomer

<sup>☆</sup> See Ref. 1.

<sup>\*</sup>Corresponding author. Tel.: +1 301 496 3569; fax: +1 301 402 0859; e-mail: kpn@helix.nih.gov

Scheme 1. Reagents and conditions: (a) NIS, AgOTf, methyl 6-hydroxyhexanoate,  $CH_2Cl_2$ , rt, 1 h; (b) NaOMe, MeOH, rt, 2 h; (c) NIS, AgOTf,  $CH_2Cl_2$ , rt, 1 h; (d) AgOTf, 2,6-di-t-Bu-Me-pyridine,  $CH_2Cl_2$ , rt, 1 h; (e)  $Cl_2$ ,  $CH_2Cl_2$ , rt, 5 min; (f) NIS, AgOTf,  $CH_2Cl_2$ , rt, 1 h; (g) 3-hydroxy-3-methylbutyric acid, HATU,  $CH_2Cl_2$ , rt, 1 h; (h) MeI,  $Ag_2O$ ,  $Me_2S$ , 1,2-dimethoxyethane, rt, 24 h; (i)  $H_2S$ , 3:1 pyridine–water, rt, 24 h; (j)  $H_2$ , Pd/C, P

Scheme 2. Reagents and conditions: (a) BrCH<sub>2</sub>COBr, TMU, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C  $\rightarrow$  rt, 24 h; (b) Ac<sub>2</sub>O, H<sub>2</sub>SO<sub>4</sub>, AcOH, rt, 1 h; (c) EtSH, BF<sub>3</sub>·Et<sub>2</sub>O, CH<sub>2</sub>Cl<sub>2</sub>, rt, 2 h.

Table 1. Comparison of  $^1H$  chemical shifts  $^a$  for tetrasaccharide 20 with data reported in Ref. 1 for 1

Ring	H-1	H-2	H-3	H-4	H-5	H-6
A	4.577	2.845	3.281	3.402	3.275	1.074
	4.582	2.850	3.301	3.401	3.278	1.082
В	4.857	3.872	3.745	3.402	3.636	1.132
	4.876	3.880	3.760	3.404	3.655	1.142
C	4.842	3.775	3.609	3.325	3.487	1.101
	4.848	3.783	3.592	3.339	3.504	1.103
D	4.566	3.634	3.530	3.155	3.373	1.134
	4.870	3.601	3.615	3.144	3.592	1.114
	NH	$CH_2$	CH <sub>3</sub>	OCH <sub>3</sub>		
Other <sup>b</sup>	7.706	2.204	1.155, 1.143	3.519		
	7.754	2.211	1.160, 1.148	3.526		

<sup>&</sup>lt;sup>a</sup> Spectra taken at 600 MHz for a solution in DMSO-d<sub>6</sub> at 25 °C. Data in the second row for each ring were taken from Ref. 1.

of 1 in the same solvent. The minute differences in chemical shifts are due to the presence of the linker substituent at *O*-1 of the D-rhamnose residue. This confirms, by chemical synthesis, the structure assigned to the material isolated from the natural source.

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## Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bmcl.2005.10.056.

 $<sup>^{\</sup>rm b}$   $\delta_{\rm OH}$  (disappear on deuteration): 4.995, 4.945, 4.885, 4.875, 4.831, 4.779, 4.718, 4.534. Not reported in Ref. 1.

Table 2. Comparison of <sup>13</sup>C chemical shifts for tetrasaccharide 20 with data reported in Ref. 1 of 1

Ring	C-1	C-2	C-3	C-4	C-5	C-6
A	103.50; J <sub>C.H</sub> 160.7	84.36	72.77	56.42	70.34	18.18
	103.4; J <sub>C.H</sub> 161	84.3	72.7	56.4	70.3	18.1
В	101.71; <i>J</i> <sub>C,H</sub> 171.3	69.67	79.48	71.65	68.31	17.73
	101.6; J <sub>C.H</sub> 171	69.6	79.4	71.6	68.2	17.6
C	101.76; J <sub>C.H</sub> 170.6	69.97	76.28	71.37	69.04	17.81
	101.6; J <sub>C,H</sub> 170	69.9	76.2	71.4	68.8	17.9
D	98.62; J <sub>C,H</sub> 167.6	76.66	70.56	72.22	68.58	18.02
	92.8; J <sub>C,H</sub> 166	77.7	70.1	72.6	67.7	17.9
	CO	$CH_2$	CH <sub>3</sub>	$C_{quat}$	$OCH_3$	
Other	171.45	48.67	29.58, 29.48	68.71	60.16	
	171.4	48.6	29.5, 29.4	Not reported	59.9	

<sup>&</sup>lt;sup>a</sup> Spectra taken at 150 MHz for a solution in DMSO-d<sub>6</sub> at 25 °C. Data in the second row for each ring were taken from Ref. 1.

## References and notes

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- 13. All reactions were monitored by TLC on silica gel coated glass slides, and the structure of products was verified by MS and NMR spectra. All new compounds produced correct analytical figures by combustion analysis, except for compounds 7, 13, 17, and 20. Copies of NMR spectra of the four compounds are available as Supplementary data.

Compound 3: 83%;  $[\alpha]_D - 6^\circ$  (c 1, CHCl<sub>3</sub>);  $\delta_{H-1}$  5.35,  $J_{1,2}$ 1.8 Hz;  $\delta_{C-1}$  97.60.

Compound 4: 96%;  $[\alpha]_D$  –39° (c 0.9, CHCl<sub>3</sub>);  $\delta_{H-1}$  4.78,  $J_{1,2}$ 1.6 Hz;  $\delta_{\text{C-1}}$  98.83.

Compound 7: 88%;  $\delta_{H-1}$  5.98,  $J_{1,2}$  1.6 Hz;  $\delta_{C-1}$  90.97. Compound 8: 63%;  $[\alpha]_D$  –42° (c 0.4, CHCl<sub>3</sub>);  $\delta_{H^{-1}^C}$  5.10,  $J_{1,2}$ 1.6 Hz,  $\delta_{\text{H-1}^{\text{D}}}$  4.67,  $J_{1,2}$  not determined due to overlap of signals;  $\delta_{C^{-1}^{C}}$  99.16,  $\delta_{C^{-1}^{D}}$  98.79.

Compound **9**: 94%;  $[\alpha]_D$  -4.6° (*c* 0.9, CHCl<sub>3</sub>);  $\delta_{H^{-1}}^B$  5.16,  $J_{1,2}$  1.7 Hz;  $\delta_{\text{H-1}^{\text{C}}}$  5.11,  $J_{1,2}$  1.9 Hz,  $\delta_{\text{H-1}^{\text{D}}}$  4.68,  $J_{1,2}$  1.8 Hz,

 $\delta_{\text{C-1}^{\text{B}}}$  99.25,  $\delta_{\text{C-1}^{\text{C}}}$  98.98,  $\delta_{\text{C-1}^{\text{D}}}$  98.87. Compound **10**: 85%; [ $\alpha$ ]<sub>D</sub> -6.6° (c 0.6, CHCl<sub>3</sub>);  $\delta_{\text{H-1}^{\text{B}}}$  5.21 (s),  $\delta_{\text{H-1}^{\text{C}}}$  5.11,  $J_{1,2}$  1.9 Hz,  $\delta_{\text{H-1}^{\text{D}}}$  4.70,  $J_{1,2}$  1.8 Hz;  $\delta_{\text{C-1}^{\text{B}}}$ 98.34, $\delta_{\text{C-1}^{\text{C}}}$  98.93,  $\delta_{\text{C-1}^{\text{D}}}$  98.82. Compound **12**: 90%; [ $\alpha$ ]<sub>D</sub> +204° (c 0.5, CHCl<sub>3</sub>);  $\delta_{\text{H-1}}$  4.86,

 $J_{1,2} \sim 3.7 \text{ Hz}; \delta_{\text{C-1}} 96.41.$ 

Compound 13: 93%;  $\delta_{\text{H-}1\alpha}$  6.24,  $J_{1,2}$  3.7 Hz,  $\delta_{\text{H-}1\beta}$  5.58,  $J_{1,2}$ 8.4 Hz;  $\delta_{C-1\alpha}$  89.04,  $\delta_{C-1\beta}$  91.41.

Compound 14: β anomer, 62%; mp 82.0-82.5 °C (from EtOH);  $[\alpha]_D$  +60° (c 0.5, CHCl<sub>3</sub>);  $\delta_{H-1}$  4.34,  $J_{1,2}$  10.0 Hz;  $\delta_{C-1}$ 82.82; the corresponding  $\alpha$ -anomer, mp 31–32 °C (from hexane),  $[\alpha]_D + 263^\circ$  (c 0.9, CHCl<sub>3</sub>); ( $\delta_{H-1}$  5.57,  $J_{1,2}$  5.4 Hz;  $\delta_{C-1}$  81.27) was formed in 23%, total yield of thioglycosidation, 85%.

Compound **15**: 84%;  $[\alpha]_D + 2.3^{\circ}$  (c 0.7, CHCl<sub>3</sub>);  $\delta_{H^{-1}^A}$  $\sim$ 4.69,  $J_{1,2} \sim$ 8.0 Hz,  $\delta_{\text{H-1}^{\text{B}}}$  5.09 (s);  $\delta_{\text{H-1}^{\text{C}}}$  5.10,  $J_{1,2}$  1.9 Hz;  $\delta_{\text{H-1}^{\text{D}}} \sim$ 4.66,  $J_{1,2}$  not determined due to overlap;  $\delta_{\text{C-1}^{\text{A}}}$  $\begin{array}{l} {}^{\text{H-I}}_{100.62,\,\delta_{\text{C-I}^{\text{B}}}} \, 100.22,\,\delta_{\text{C-I}^{\text{C}}} \, 98.75,\,\delta_{\text{C-I}^{\text{D}}} \, 98.87. \\ \text{Compound} \, \textbf{16}; 93\%; [a]_{\text{D}} \, + \, 10.4^{\circ} (c\,0.5,\text{CHCl}_3); \delta_{\text{H-I}^{\text{A}}} \, 4.39, \\ \end{array}$ 

 $J_{1,2}$  7.7 Hz,  $\delta_{\text{H-1}^{\text{B}}}$  5.08 (s),  $\delta_{\text{H-1}^{\text{C}}}$  5.11,  $J_{1,2}$  2.0 Hz,  $\delta_{\text{H-1}^{\text{D}}}$  4.67,  $J_{1,2}$  1.7 Hz;  $\delta_{\text{C-1}^{\text{A}}}$  103.70,  $\delta_{\text{C-1}^{\text{B}}}$  99.17,  $\delta_{\text{C-1}^{\text{C}}}$  98.70,  $\delta_{\text{C-1}^{\text{D}}}$ 

Compound 17: 96%;  $[\alpha]_D + 2^\circ$  (c 1, CHCl<sub>3</sub>);  $\delta_{H^{-1}^A}$  4.61,  $\begin{array}{l} J_{1,2} \ 8.0 \ \mathrm{Hz}, \ \delta_{\mathrm{H}\text{-}1^{\mathrm{B}}} \ 5.11 \ (\mathrm{s}); \ \delta_{\mathrm{H}\text{-}1^{\mathrm{C}}} \ 5.08, \ J_{1,2} \ 1.8 \ \mathrm{Hz}; \ \delta_{\mathrm{H}\text{-}1^{\mathrm{D}}} \\ 4.66, \ J_{1,2} \ 1.8 \ \mathrm{Hz}; \ \delta_{\mathrm{C}\text{-}1^{\mathrm{A}}} \ 103.52, \ \delta_{\mathrm{C}\text{-}1^{\mathrm{B}}} \ 100.26, \ \delta_{\mathrm{C}\text{-}1^{\mathrm{C}}} \ 98.96, \end{array}$  $\delta_{\text{C-1}^{\text{D}}}$  98.90.

Compound **18**: 88%;  $\delta_{\text{H-1}^{\Lambda}}$  4.70,  $J_{1,2}$  7.7 Hz,  $\delta_{\text{H-1}^{B}}$  5.12 (s),  $\delta_{\text{H-1}^{C}}$  5.08,  $J_{1,2}$  1.9 Hz,  $\delta_{\text{H-1}^{D}}$  ~4.66,  $J_{1,2}$  not determined due to overlap;  $\delta_{\text{C-1}^{\Lambda}}$  103.82,  $\delta_{\text{C-1}^{B}}$  100.37,  $\delta_{\text{C-1}^{\text{C}}}$  99.03,  $\delta_{\text{C-1}^{\text{D}}}$  98.89. Compound **19**: 97%; [a]<sub>D</sub> –26° (*c* 0.5, CHCl<sub>3</sub>);  $\delta_{\text{H-1}^{\text{A}}}$  4.62,

 $\begin{array}{l} J_{1,2} \ 8.0 \ \mathrm{Hz}, \ \delta_{\mathrm{H^{-1}}^{\mathrm{B}}} \ 5.09 \ (\mathrm{s}), \ \delta_{\mathrm{H^{-1}}^{\mathrm{C}}} \ 5.07, \ J_{1,2} \ 2.0 \ \mathrm{Hz}, \ \delta_{\mathrm{H^{-1}}^{\mathrm{D}}} \\ \sim & 4.66, \ \sim & 1.9 \ \mathrm{Hz}; \delta_{\mathrm{C^{-1}}^{\mathrm{A}}} \ 103.69, \ \delta_{\mathrm{C^{-1}}^{\mathrm{B}}} \ 100.34, \ \delta_{\mathrm{C^{-1}}^{\mathrm{C}}} \ 98.94, \end{array}$  $\delta_{\text{C-1}^{\text{D}}}$ 98.86.

Compound **20**: 83%;  $[\alpha]_D$  –52.6° (c 0.5, H<sub>2</sub>O).